

Sampling and Source Tracking of *Cryptosporidium* spp. Oocysts

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Ultimate Goal of Source Tracking

- Identify important watershed sources of *Cryptosporidium* spp. oocysts
- Develop watershed management policies to prevent *Cryptosporidium* spp. contamination of surface waters

Methods

Surface Water Filtration Fecal Sample Collection



DNA Extraction

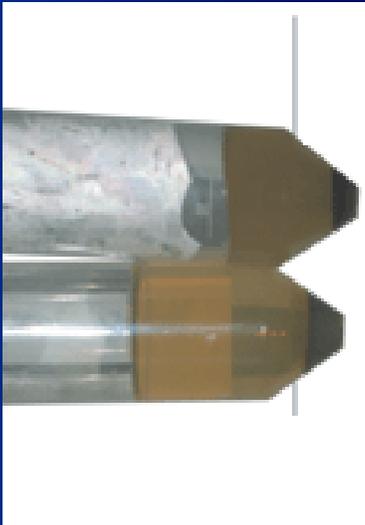
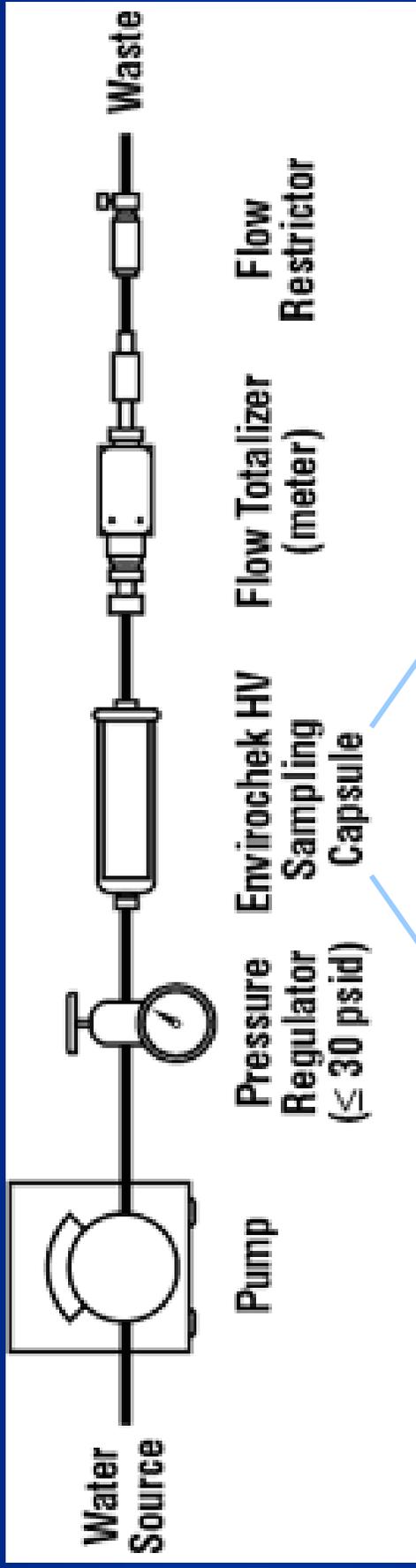


Nested PCR



Clone & Sequence

Surface Water Filtration: Gelman Envirochek Filter Capsules



Methods

Surface Water Filtration Fecal Sample Collection



Immunomagnetic Separation



DNA Extraction

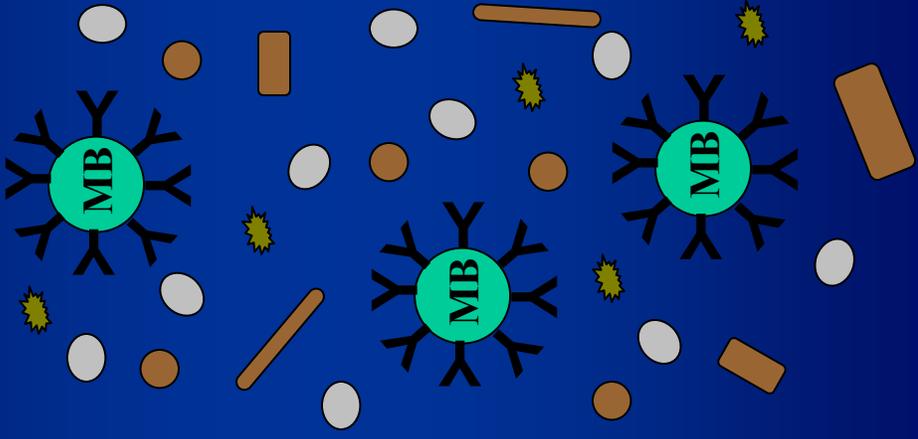


Nested PCR

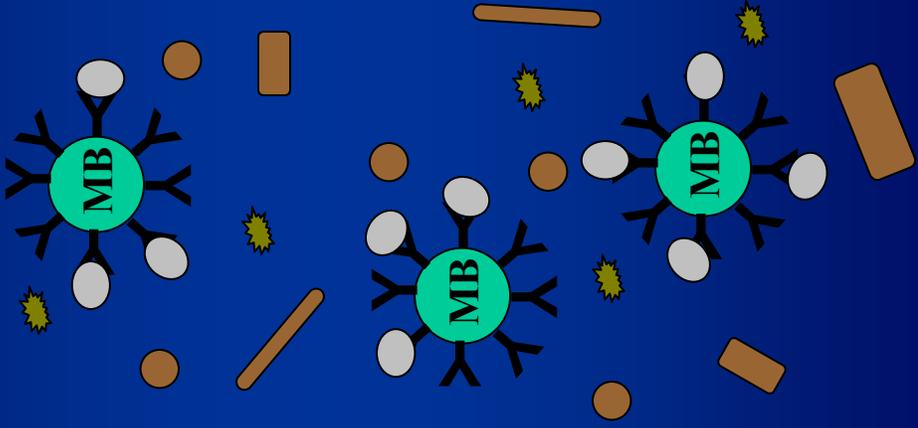


Clone & Sequence

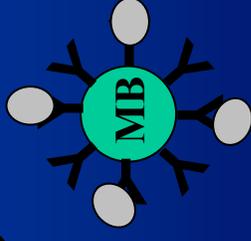
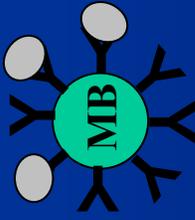
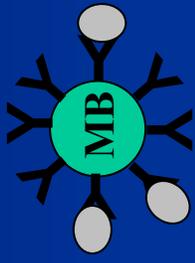
Immunomagnetic Separation



Immunomagnetic Separation



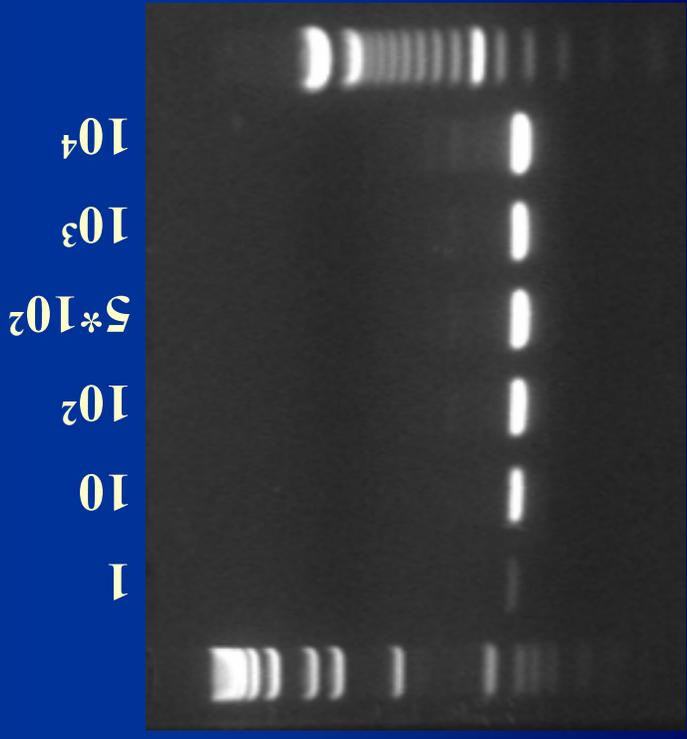
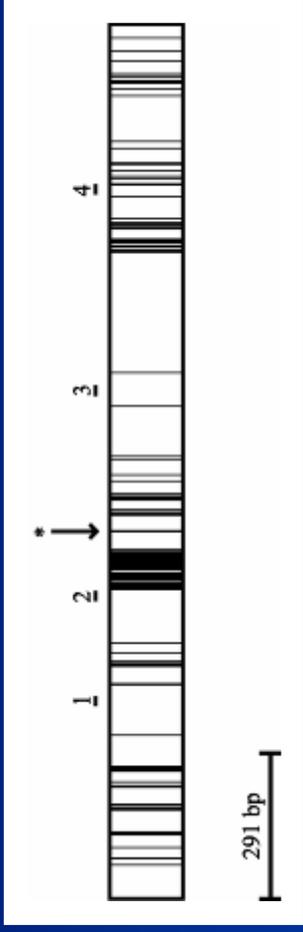
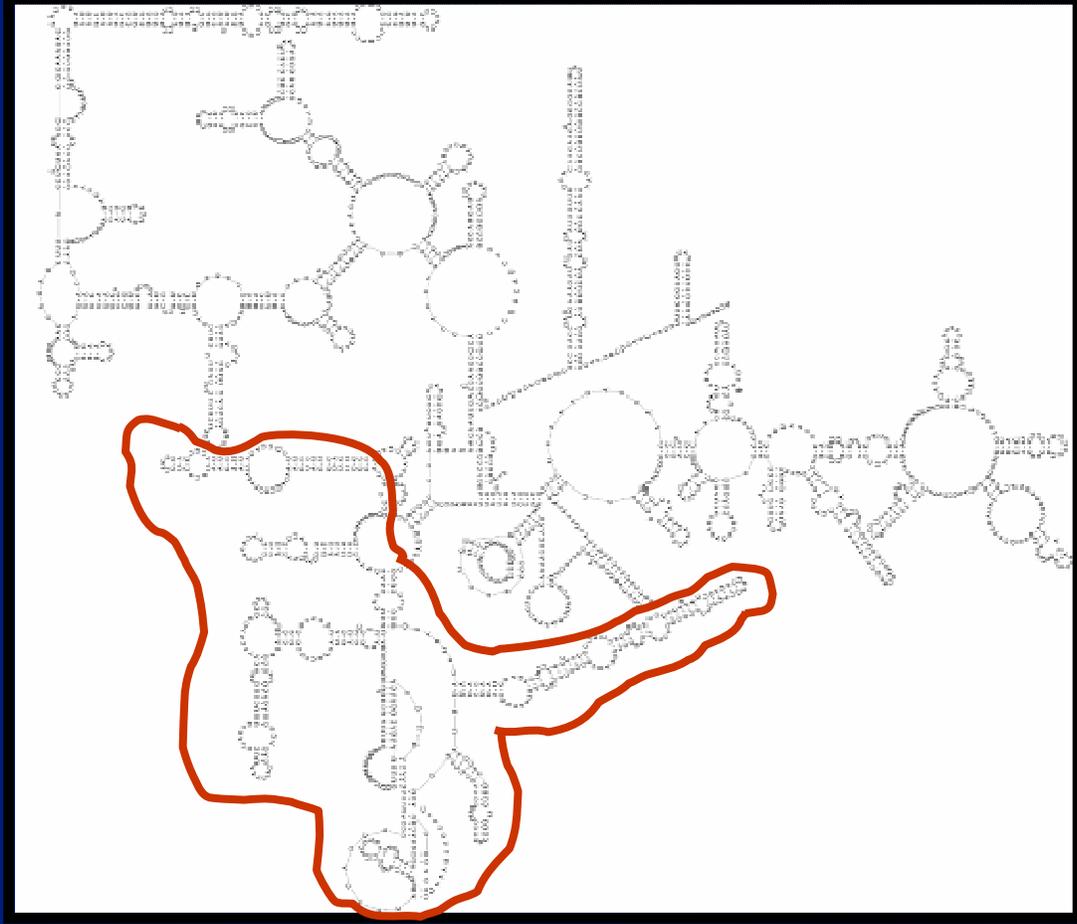
Immunomagnetic Separation



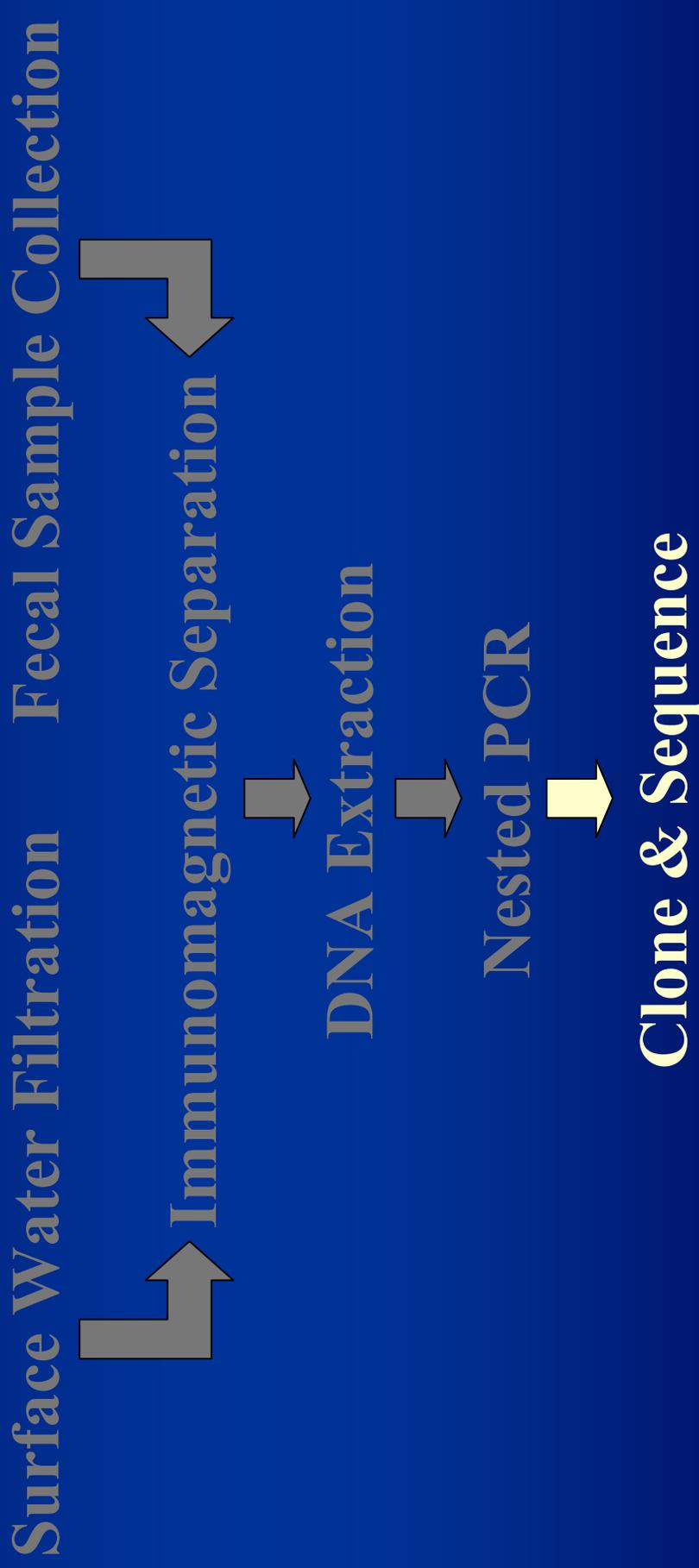
Methods



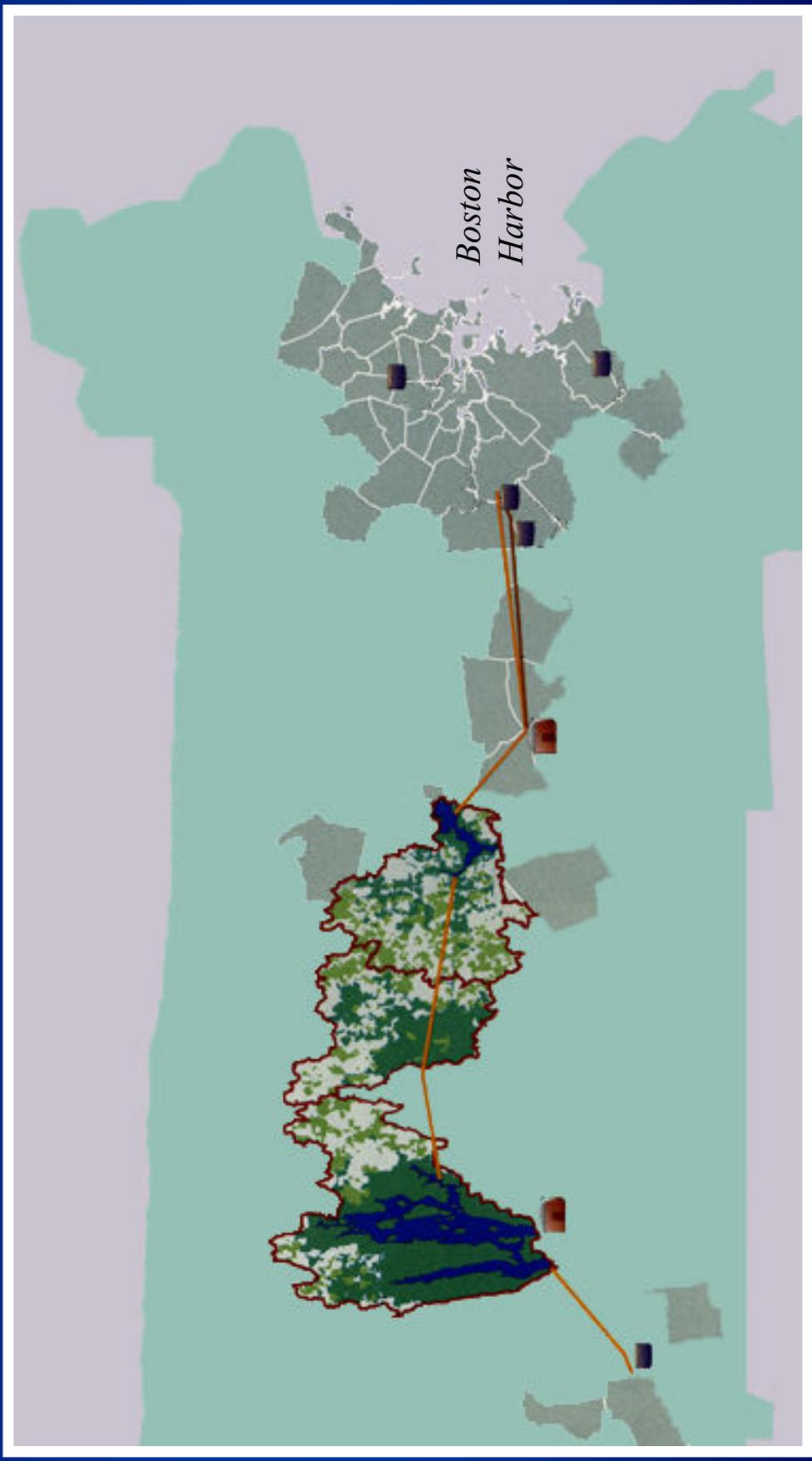
Nested PCR: 18S rRNA



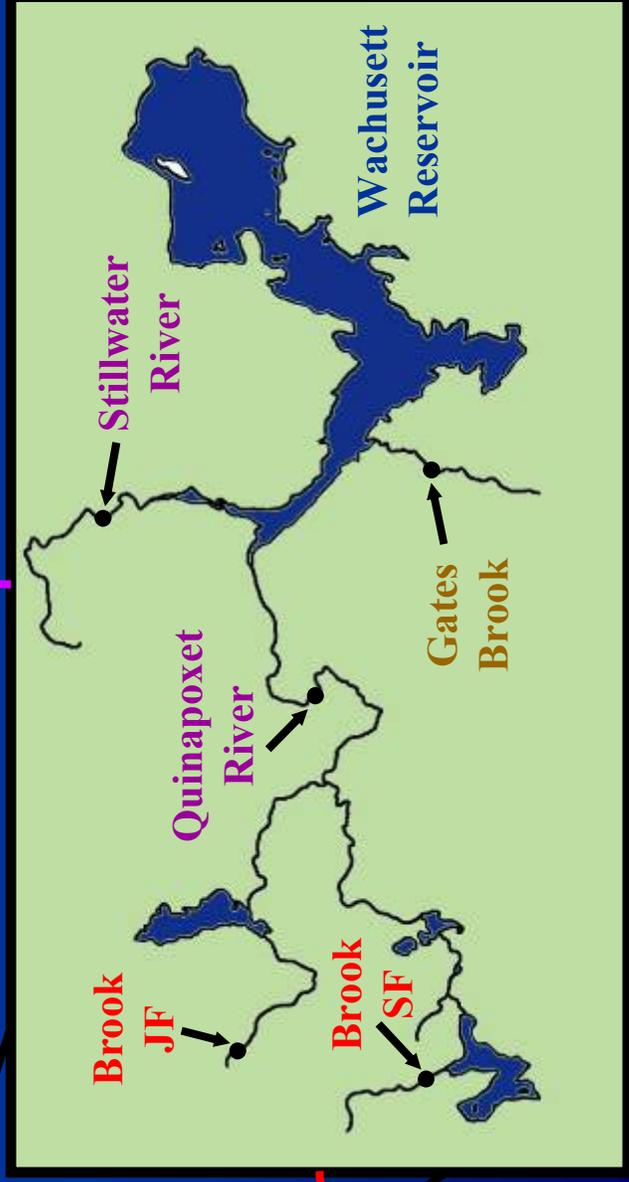
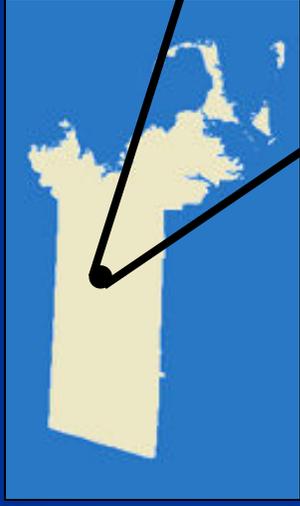
Methods



MWRA Water Supply System



Wachusett Reservoir Watershed: Surface Water Sampling Sites



Observations: Wachusett Reservoir Watershed

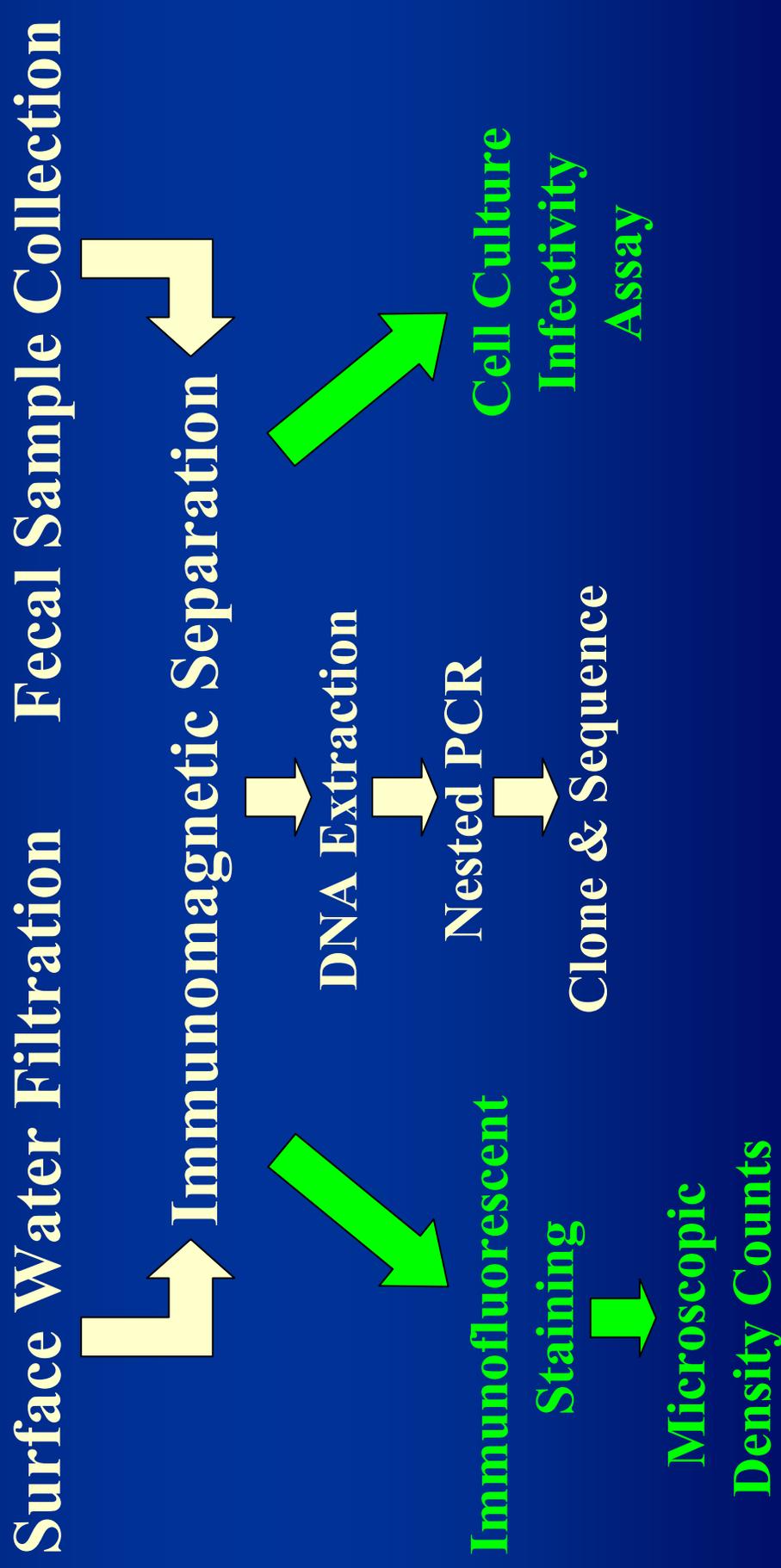
- The molecular detection method is sensitive and specific for *Cryptosporidium* spp. oocysts in water and fecal samples
- Nested PCR is required for detection of *Cryptosporidium* spp. in baseline environmental samples
- Multiple species of *Cryptosporidium* are detectable in environmental samples
- Suspected oocyst source is not always the observed source
 - Evidence of wildlife, not human, impacts on GB
 - Birds seem to be a bigger influence on JF than dairy farm activity

Method Comparison

	EPA Method 1622	Molecular Detection Method
Quantitative estimate of oocyst density	Yes	No*
Identification of oocyst infectivity	No	No*
Detection of low oocyst densities (e.g., baseline environmental samples)	No	Yes
Identification of oocyst genotype	No	Yes

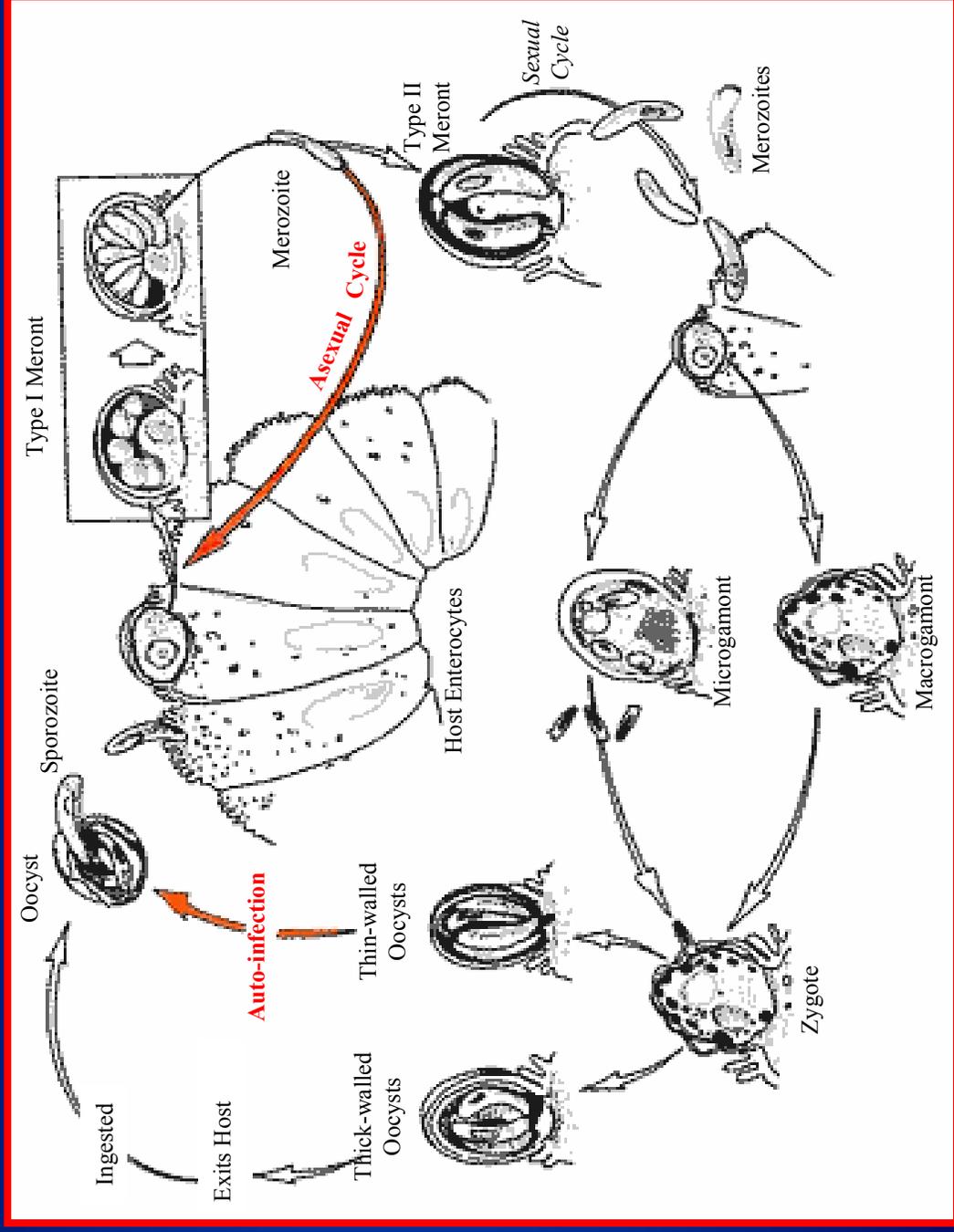
*But could be determined with method modifications

Methods



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Cryptosporidium Life Cycle



(Adapted from Current & Blagburn, 1990)