

Detection of *Cryptosporidium* by EPA Method 1623: A Summary of the Method and Method Validation

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Workshop on Developing a Plan to Identify
Cryptosporidium Sources in the Potomac River

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Objectives

- Method Evaluation Criteria
- Method 1623 Lab Validation Criteria
- Overview of EPA Method 1623
- Summary of Results with Method 1623
- Conclusions

Challenges in Detecting Environmental Parasites

- Tend to occur in low numbers
- Tend to occur intermittently
- Mix of human and animal pathogens
- Mix of viable, stressed, and dead organisms
- Debris and water chemistry

Environmental Methods and Laboratory Analysis

- Multiple analytical approaches
 - Information gathered
 - Performance efficiencies
 - Multiple analysts and labs
- Performance-based Methods
 - Similar approach, different protocols
- Different water matrices
- QC/ QA

Method Evaluation Criteria

Lindquist, et al, 1999

- Statistical Performance
 - % Recovery
 - Detection limit
 - Precision
 - Confidence intervals
 - Specificity
 - False positive rate
- Collaborative Testing

Method Evaluation Criteria, cont.

- Data Generated
 - Genus/ species
 - Viability
- Method Description
 - Completeness
 - Scope and interferences
 - Materials, supplies, and equipment
 - Data reporting, interpretation, analysis
 - Sampling
 - Range of volumes
 - Holding times and preservation
 - Managerial Criteria
 - Training, personnel, etc.

US EPA Method 1623

- Benchmark for detecting and quantifying *Giardia* and *Cryptosporidium* in water
- <http://www.epa.gov/microbes/>
- Performance-based method
- Genus level analysis
- Viability not determined
- Multi-lab validated

Spikes for Recovery Studies

- Live oocysts
 - Age (<6 weeks)
 - Enumeration method
 - Flow cytometry
 - Serial dilution
 - Time and storage effects
- Gamma-irradiated oocysts
 - EasySeed
 - Warnecke, et al, 2003
- Differentiated oocysts
 - ColorSeed
 - Francy, et al, 2004

Single Lab Validation and QA Criteria for *Cryptosporidium*

- Initial Performance and Recovery (IPR)
 - Spiked reagent water
 - Mean of 4 tests (>24% recovery, RSD 55)
- Ongoing Precision and Recovery (OPR)
 - 1 spiked reagent water/ week or 20 samples
 - >11% recovery
- Method Blanks
 - 1 with IPR and 1 weekly
- Matrix spike (MS)
 - Spiked field sample and reference sample
 - 1/ 20 field samples, over at least 12 months per water type
 - 15-118% recovery

Multi-lab Validation Criteria

- 3 labs must meet single lab's IPR, OPR, and Method blanks
- MS/ MS duplicate
 - 2 spiked field samples with reference samples
 - 1/ 20 samples, over at least 12 months per water type
 - 15-118% recovery average, RSD 30

Method 1623 Analytical Process Flow Diagram

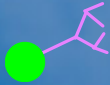


Filtration

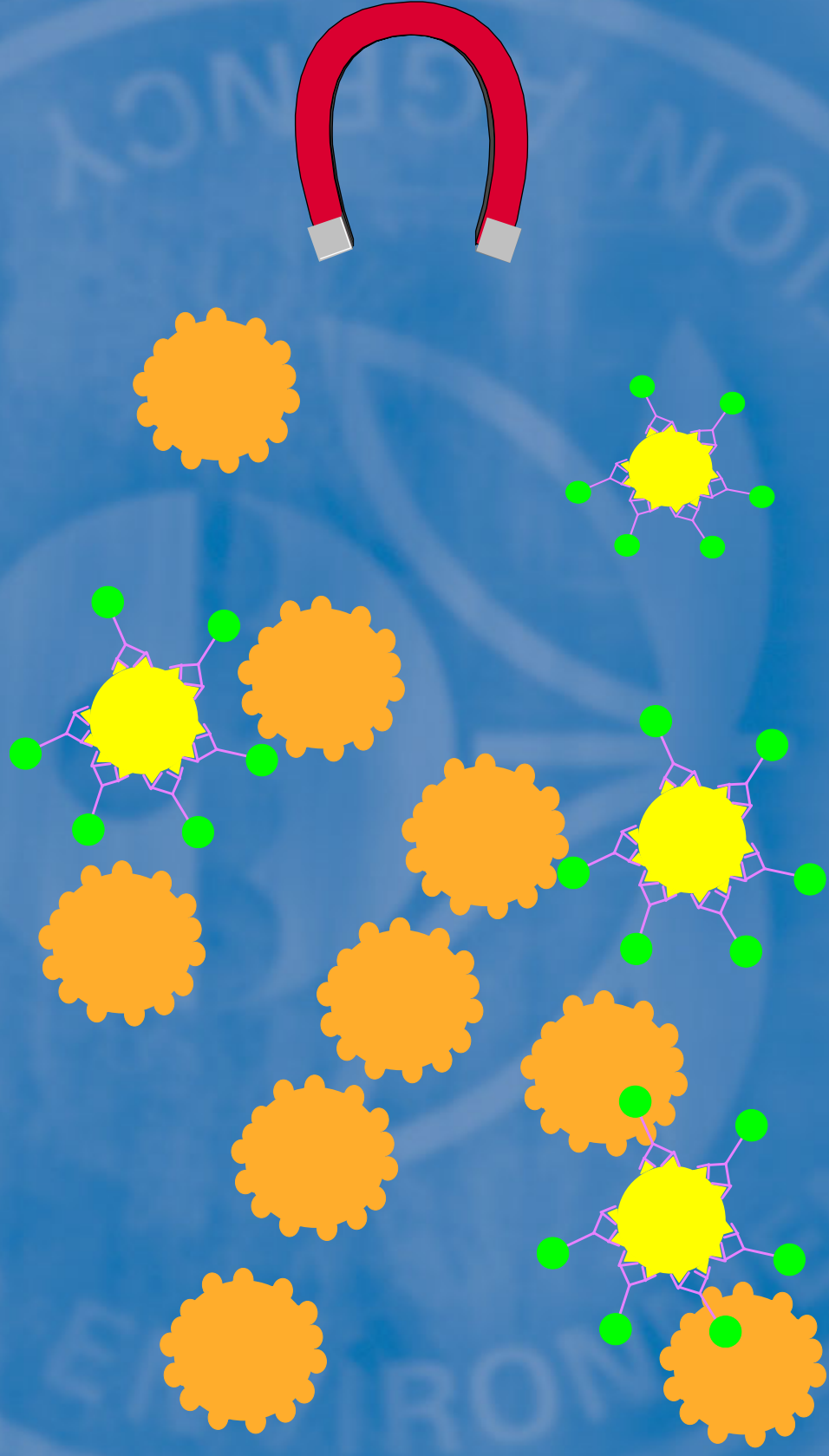
- Raw water 10-50 L
 - Finished water up to 1000 L
 - Flow rates 1 to 4 L/ min
- 4 validated filters
- Envirochek, Envirochek HV, and Cryptest all 1 μm porosity
 - Filtamax nominal 1 μm porosity

Immunomagnetic Separation (IMS)

Label with antibody
conjugated to
paramagnetic bead

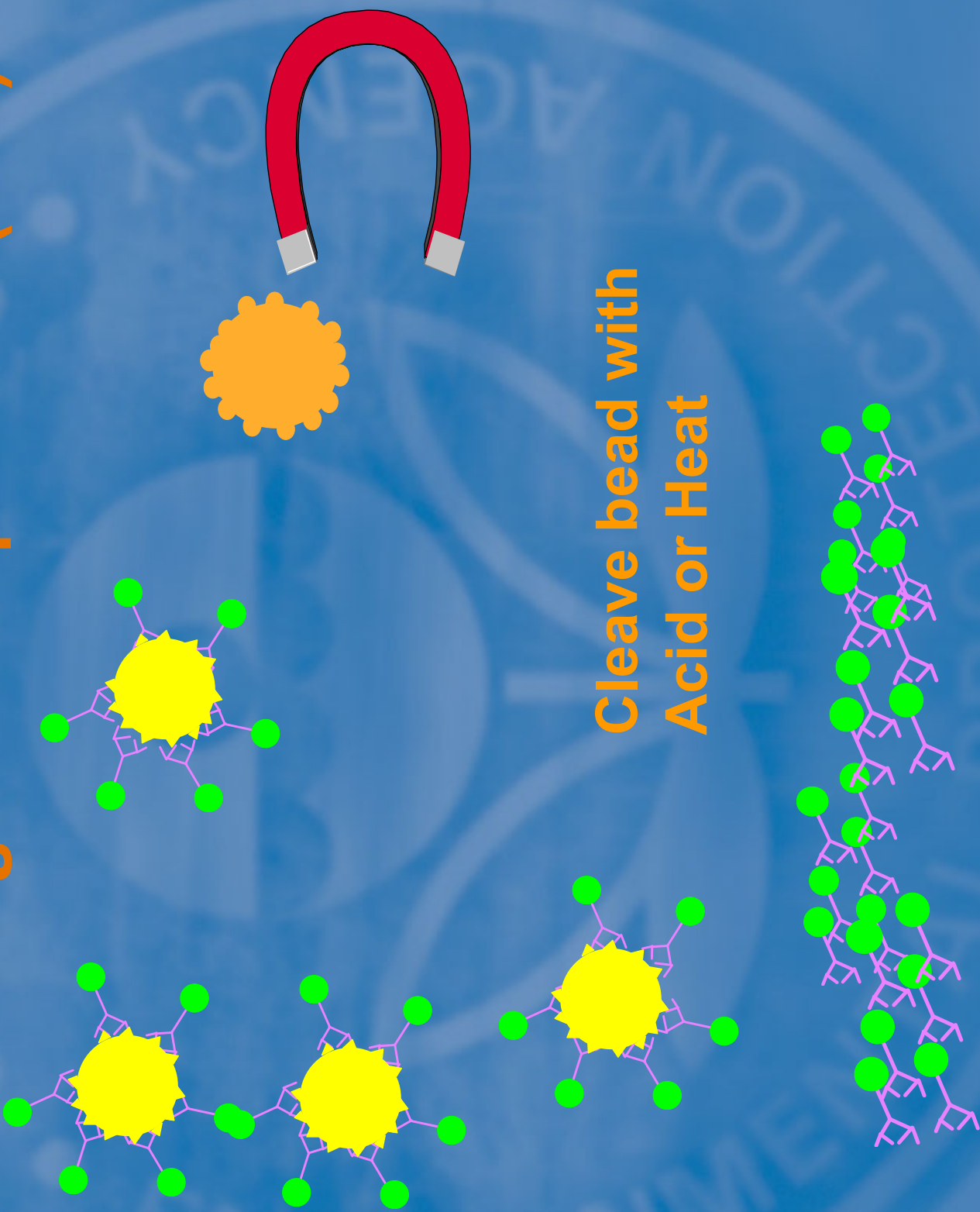


Immunomagnetic Separation (IMS)



Capture with magnetic field,
remove debris

Immunomagnetic Separation (IMS)

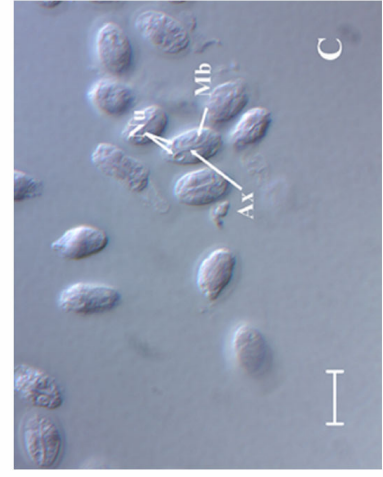
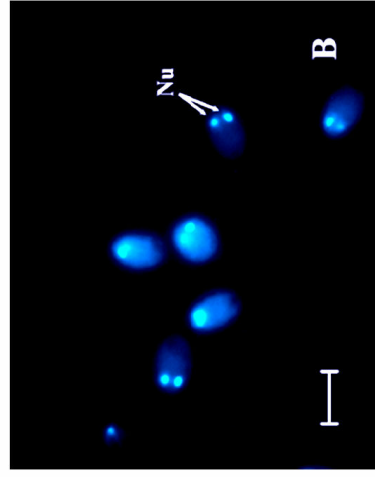
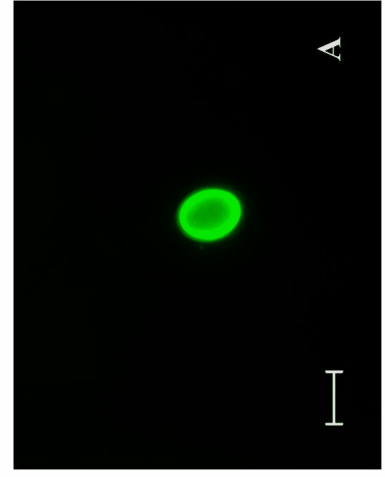


Cleave bead with
Acid or Heat

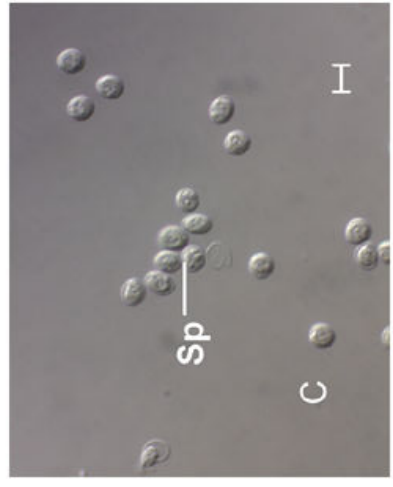
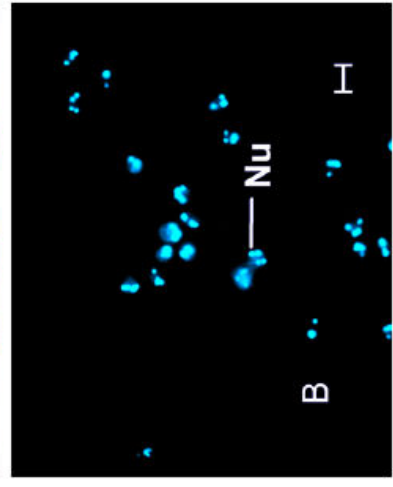
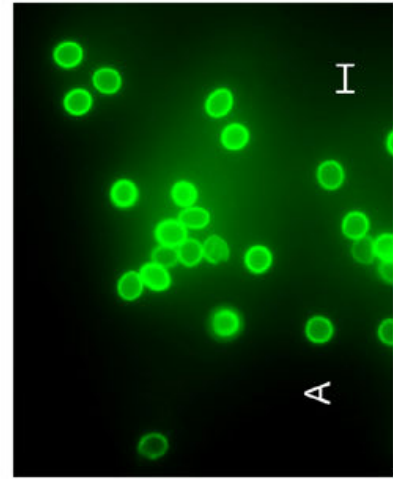
Staining and Microscopy

- Oocysts and cysts dried onto slide
- Stain with FITC conjugated monoclonal antibody (mAb) and DAPI
- Primary detection by mAb
- Confirm by detecting DAPI stained nuclei and/or internal structures

Giardia



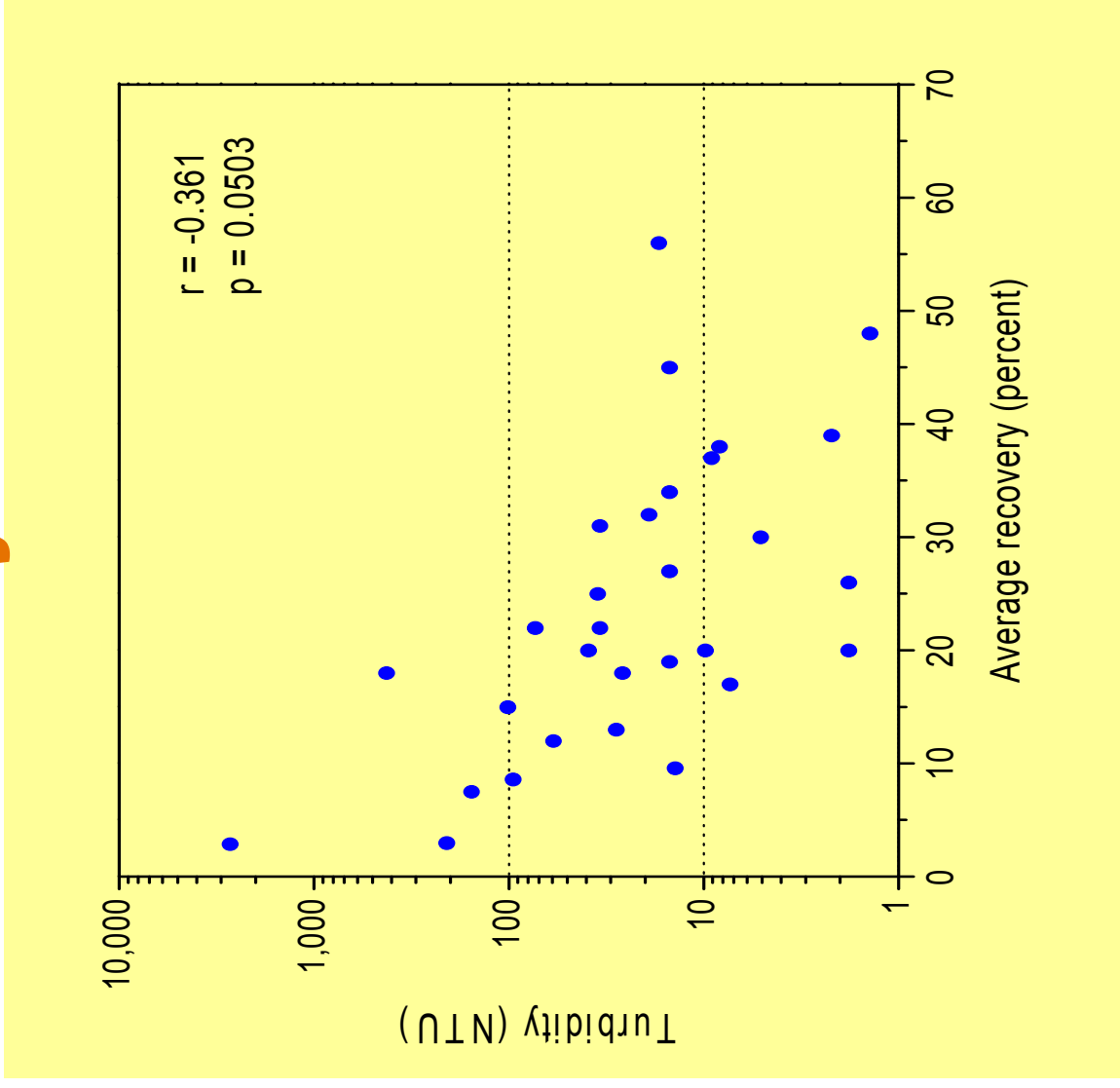
Cryptosporidium



Results from Studies Evaluating Method 1623

- Most evaluated performance in surface, ground, or drinking water.
- Adversely affected by turbidity
- Wide range of recoveries reported (0% to 80%+)
- Prone to false negative and false positive results

Turbidity Effect



Francy, et al, 2004

Conclusions

- Method 1623
 - Matrix effects
 - Variable performance
- How do you interpret results?
 - Good QA program
 - Performance criteria
- How do compare results from different labs?
 - Lab validation
 - Objective comparison criteria